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MEDICINE

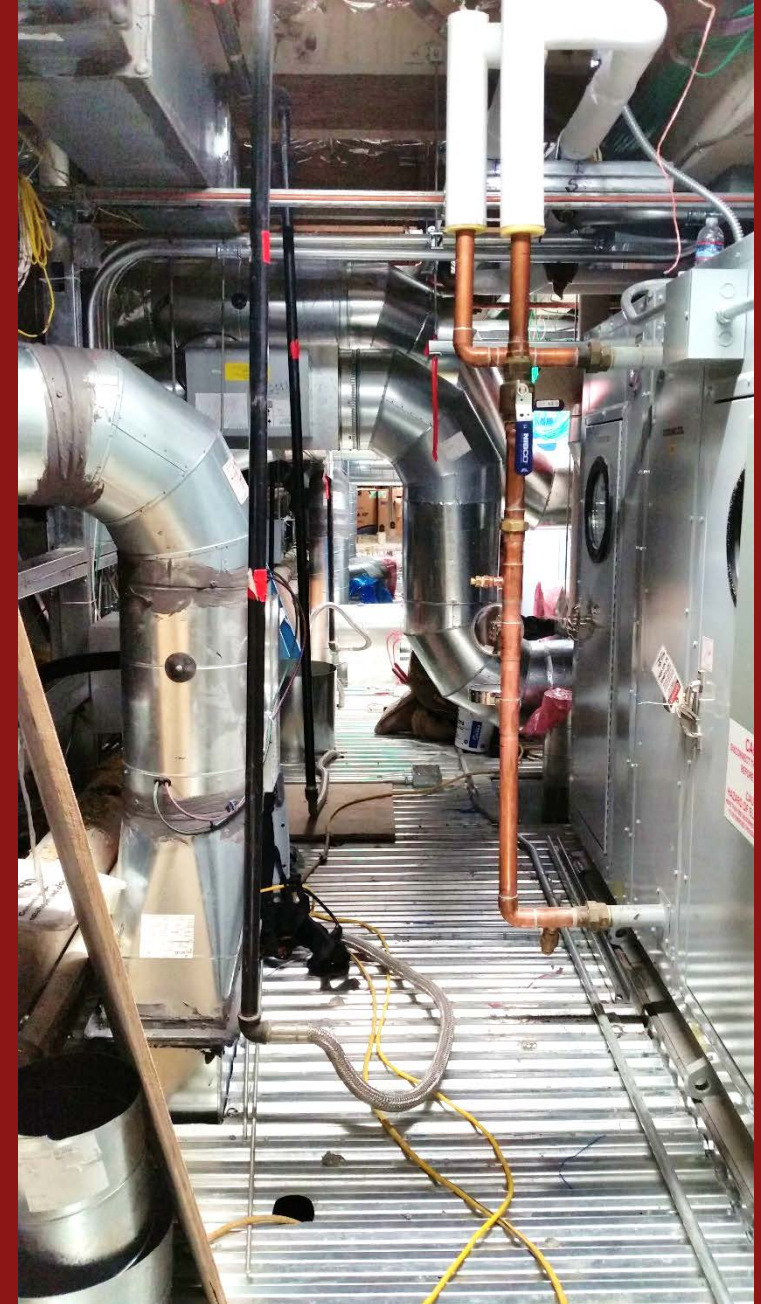
# Academic Cell and Gene Therapy Development and Manufacturing

David DiGiusto PhD

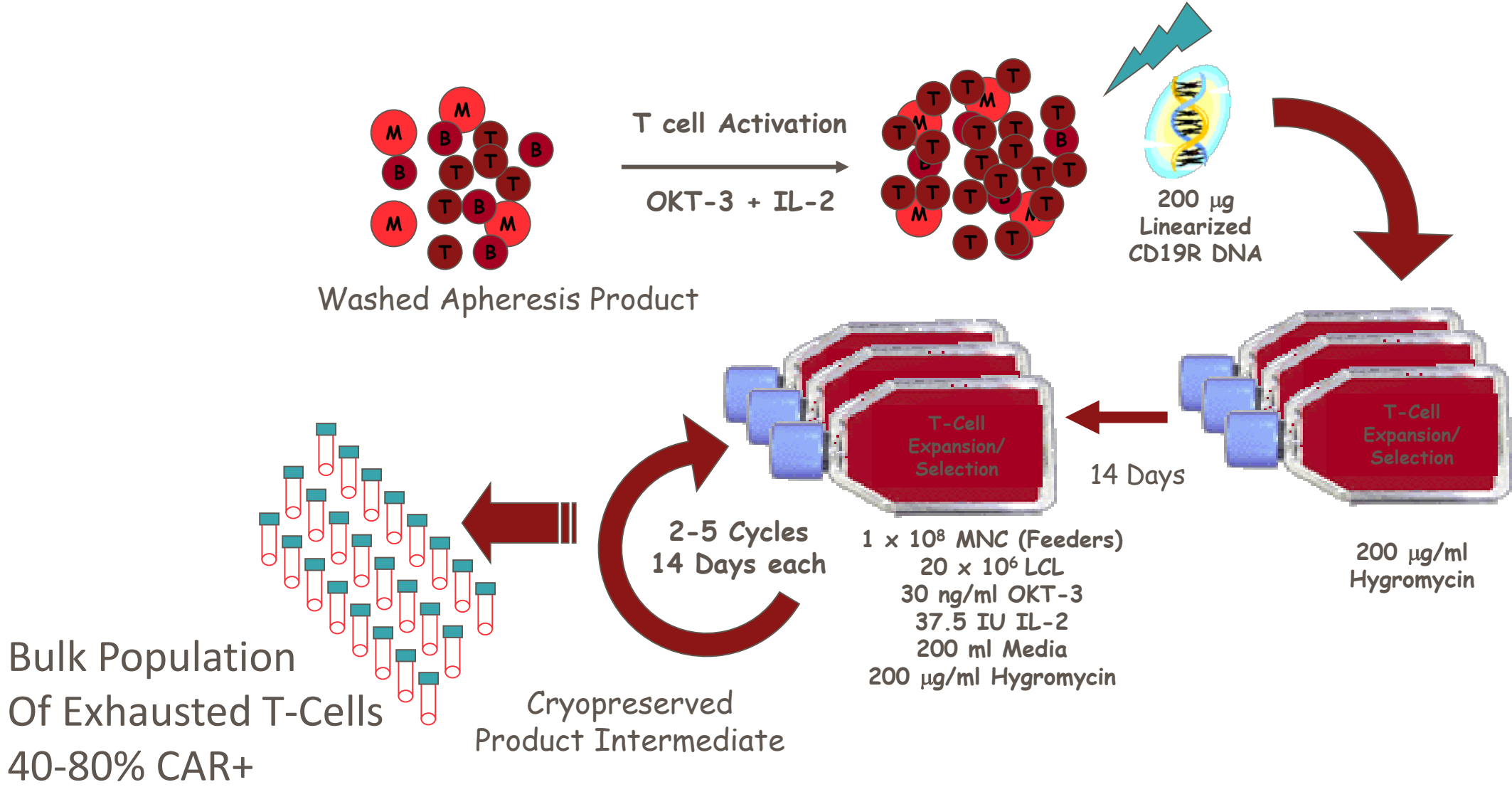
Executive Director

Stem Cells and Cellular Therapeutics Operations

Stanford University School of Medicine



# T-cell Manufacturing Process circa 2000



# Fast forward 2016 - Bi-Specific CAR-T cells

- **Process Challenges**
  - Closed system processing of Apheresis products
  - CD4/CD8 T-cell enrichment required
  - Lentiviral Transduction in closed system
  - Bioreactor culture and harvest in closed system
- **Analytical Challenges**
  - Demonstrating simultaneous expression of both CARs
  - Deciding on in process metrics
  - Rapid reliable assay for RCL detection

# Closed and (Semi) Automated Blood Cell Culture Systems

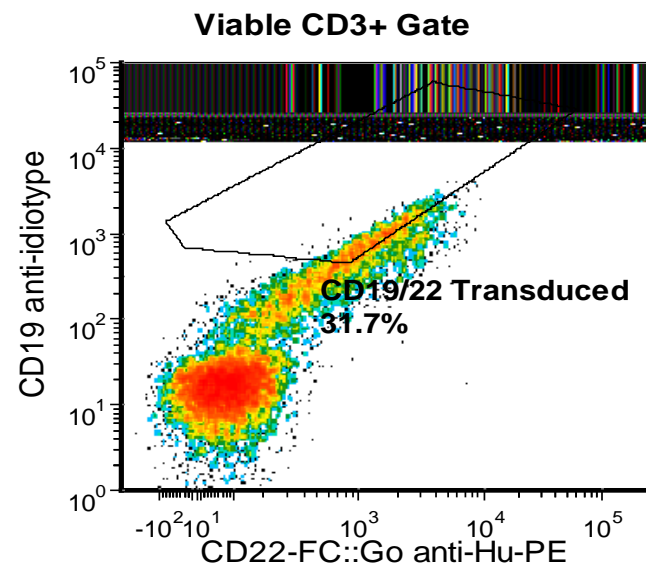
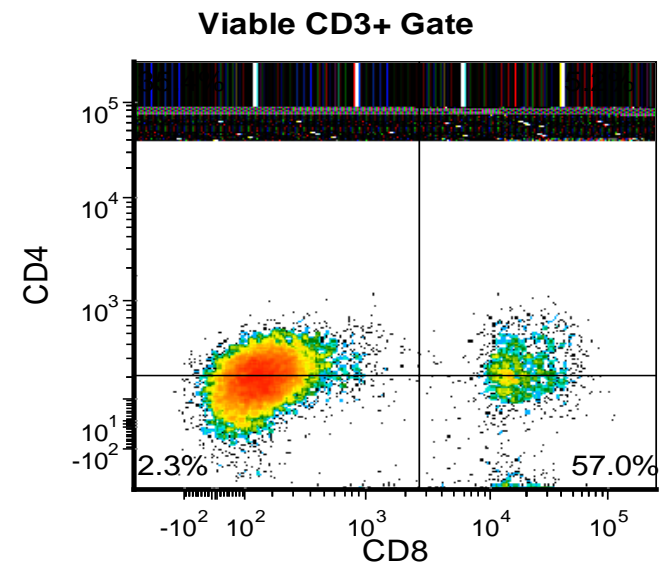
7 Day Process on Prodigy  
Apheresis Washing  
Magnetic Bead Labelling  
Target Cell Enrichment  
Cell Growth Formulation  
Cell Stimulation  
Viral Transduction  
Cell Expansion  
Cell Harvest  
Release Testing

CliniMACS Prodigy



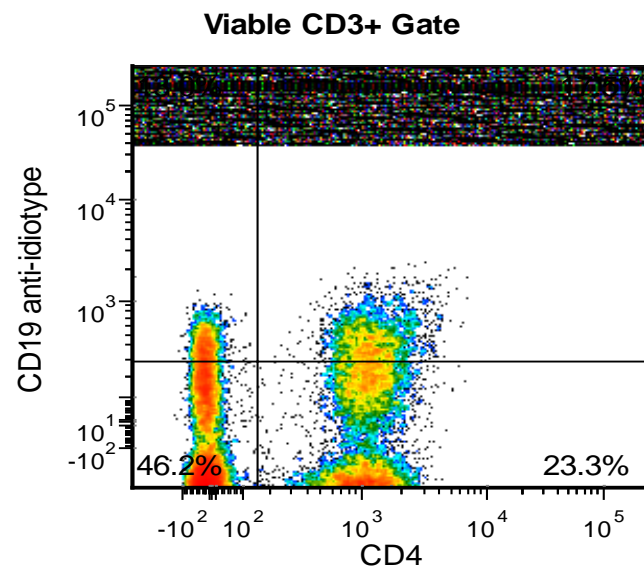
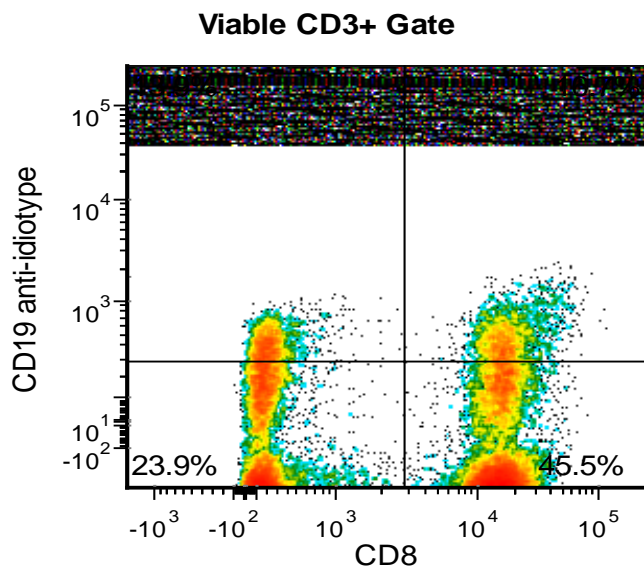
CAR T-cells

# Phenotypic Analysis of CD19/CD22 bi-Specific CAR-T



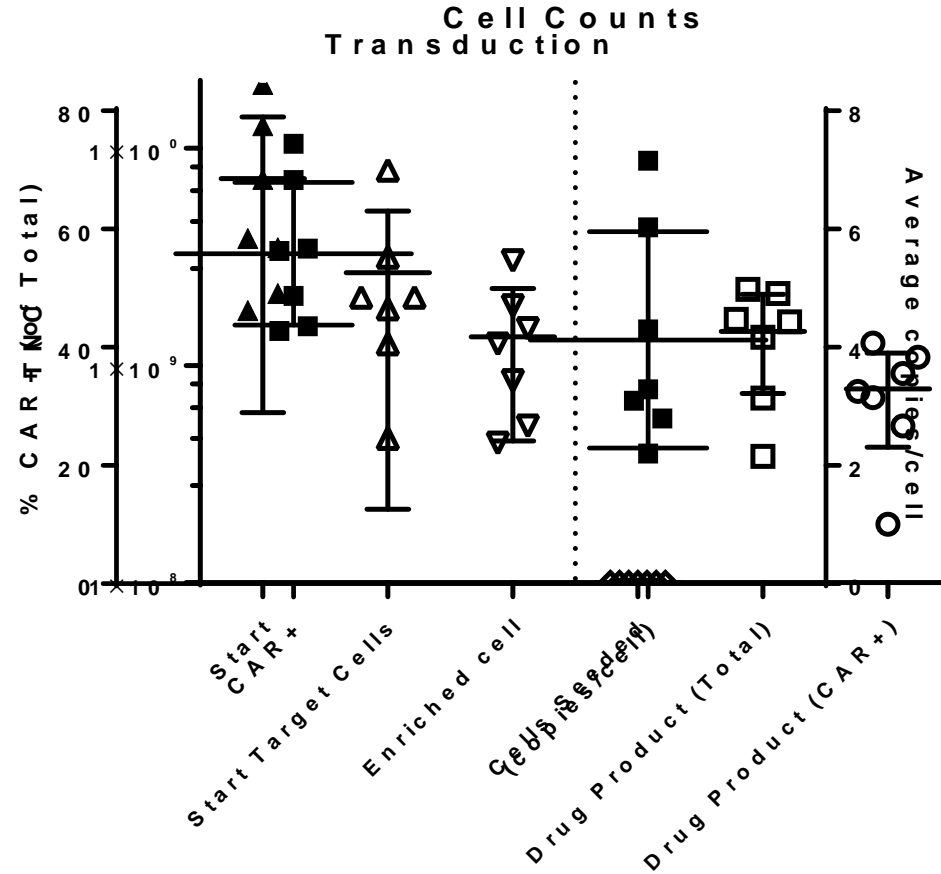
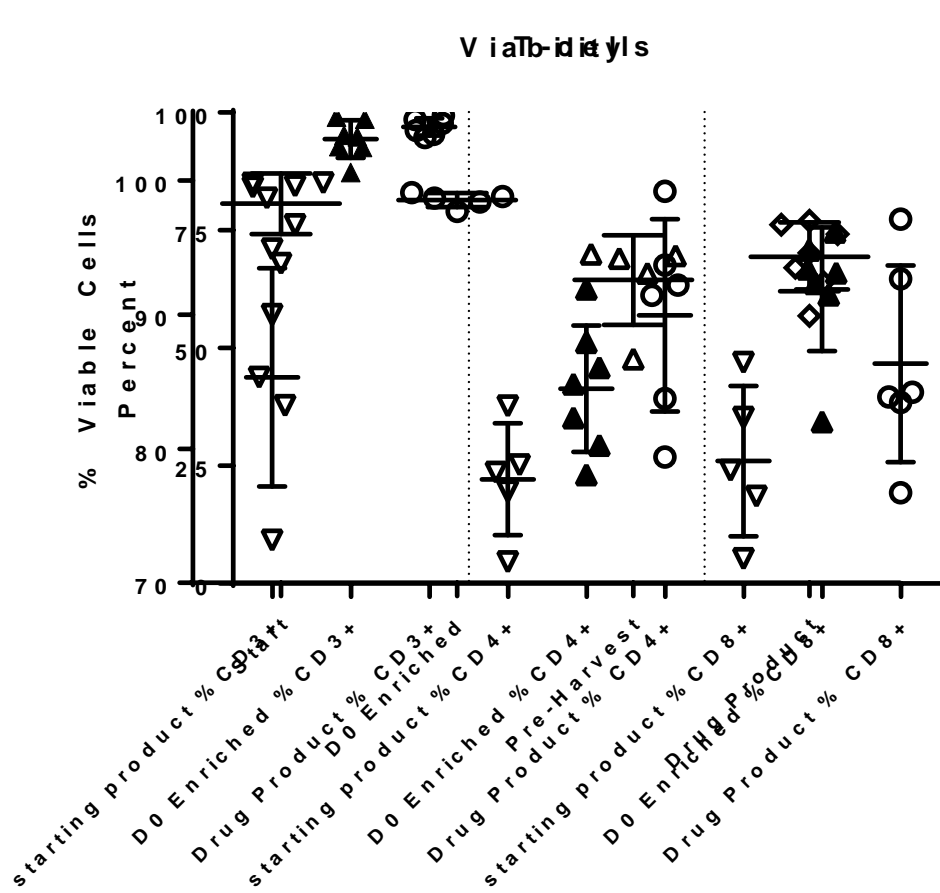
Anti-CD19 idotypic antibody to CD19 CAR  
(Laurence Cooper)

rhSiglec 2/Fc Chimera  
(CD22 on Ig stalk – R&D Systems)





# CAR-T Production Metrics



# Production summary

- Pros
  - Reproducible production of CAR-T cells
  - All cells passed all release testing
  - Semi-Closed system for production
    - Still open harvest/formulation steps performed in BSC
- Cons
  - Sole source provider for all reagents/tubing sets
  - Multiple device failures (valves, motors, tubing sets)
  - Expensive incubator for 6/7 days
    - Limits productivity in GMP suite

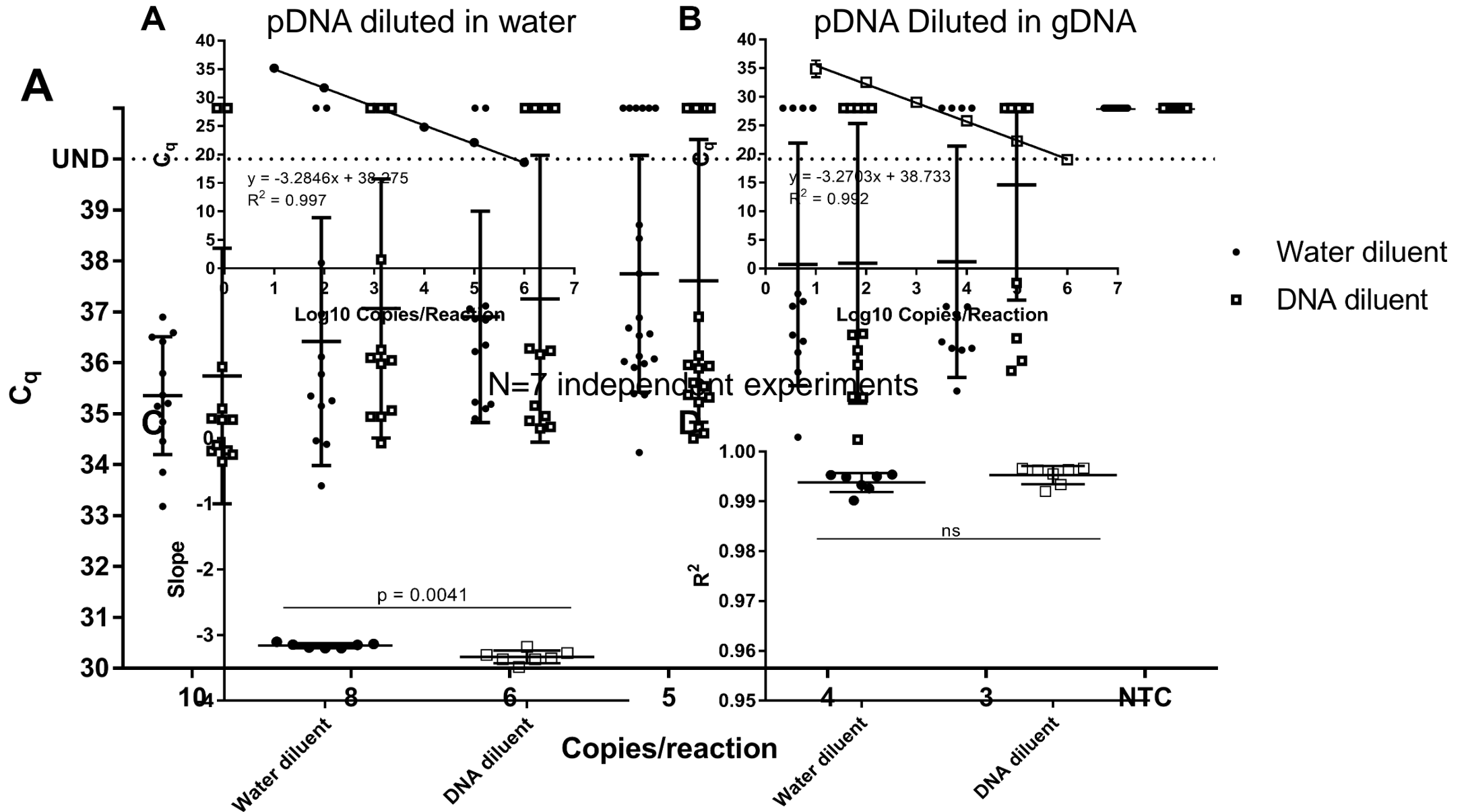
# qPCR for detection of RCL

- Using VSV-G as a target
  - All lentiviral constructs contain VSV-G envelop sequences
  - Env is on a helper plasmid and should not be contained in transduced T-cells
  - RCL could be formed in VSV-G env is transferred to T-cells
- Determine Quality of the assay (MIQE)
  - Carefully quantified plasmid DNA as Control
  - Linearity, slope are as predicted by dilutions
  - Specificity and Limit of detection is established
    - Measure frequency of false positive and false negative

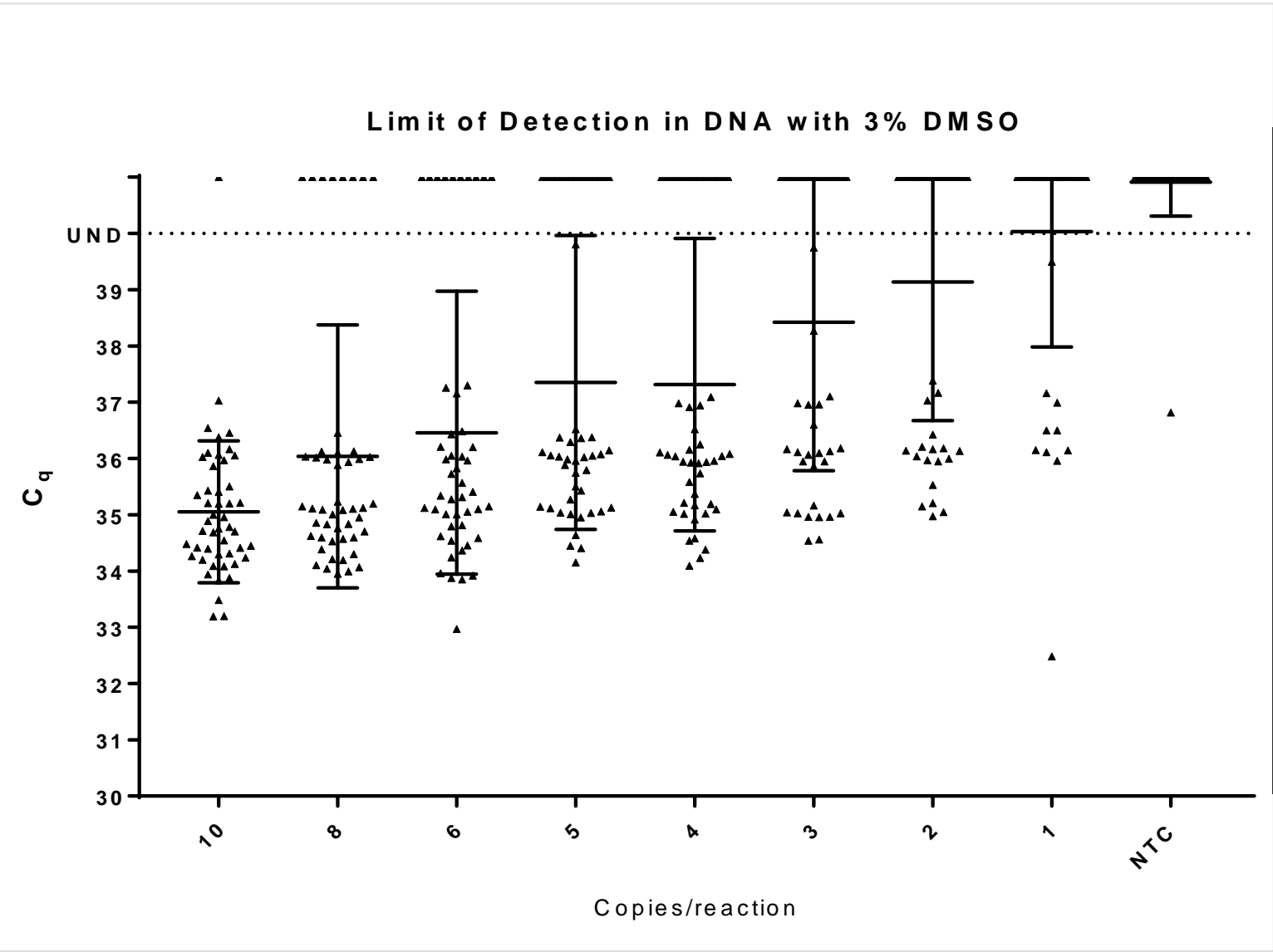
Minimum Information for Publication of Quantitative Real-Time PCR Experiments (MIQE)



# Linearity and slope of qPCR of VSV-G Plasmid



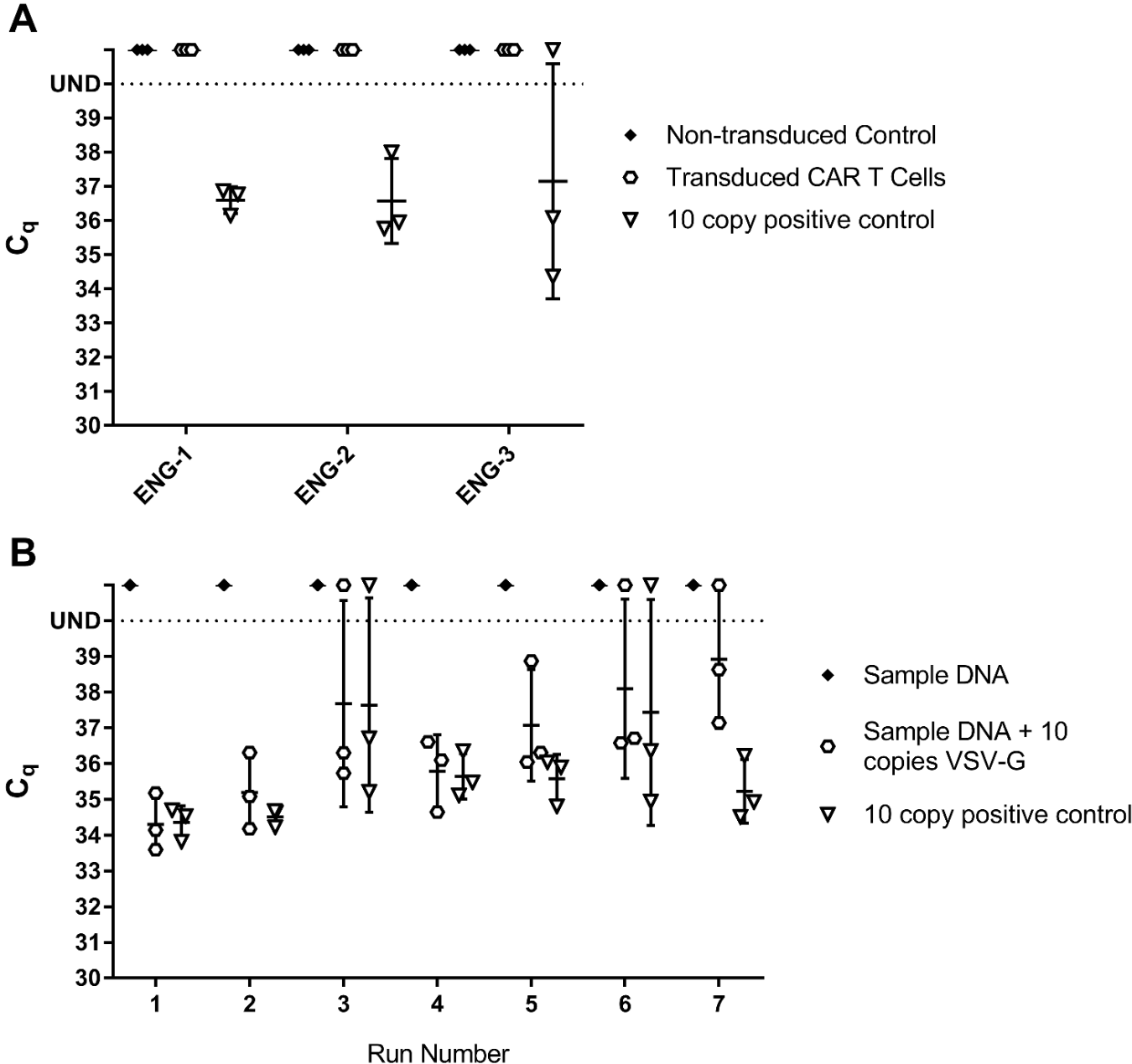
# Inclusion of 3% DMSO Improves Detection of Low Copy Number DNA



	Water Diluent	DNA diluent	DNA diluent + 3% DMSO
10 copies	100.00%	83.33%	97.92%
8 copies	83.33%	75.00%	83.33%
6 copies	83.33%	66.67%	79.17%
5 copies	76.19%	61.90%	68.75%
4 copies	66.67%	58.33%	68.75%
3 copies	66.67%	33.33%	52.08%
NTC	0.00%	0.00%	2.08%

N = 16 independent experiments in triplicate

# Pre-Clinical Qualification Runs and First 7 patient samples



# Analytical Summary

- Pros
  - Simultaneous Detection of expression of both CAR proteins
  - Reproducible, sensitive RCL assay
    - Reliable detection of 10 copies of VSV-G
    - False Negative rate <3%
    - False Positive rate ~2%
    - Low Inter-operator variability
  - All testing completed and reviewed on day of product harvest (12 hours)
- Cons
  - Requires clean, dedicated space for assays
  - Requires carefully quantified source of control DNA
  - Approximately 1/40 products may be falsely considered positive for RCL
  - Approximately 1/33 products may miss 10 copies of RCL

# Acknowledgments

- LCGM CAR-T Production Staff
  - Lindsey Skrdlant, Kerri Tate, Cindy Tudesco, Brian Fox
- LCGM Analytical Team
  - Brett Keidaisch, Mario Lorente, Randall Armstrong
- Collaborators
  - Stanford Cancer Immunology and Immunotherapy Program (**Crystal Mackall**)
  - Miltenyi Biotech Tim Waters
  - NCI – Terri Fry

**Thank You for Your Attention!**