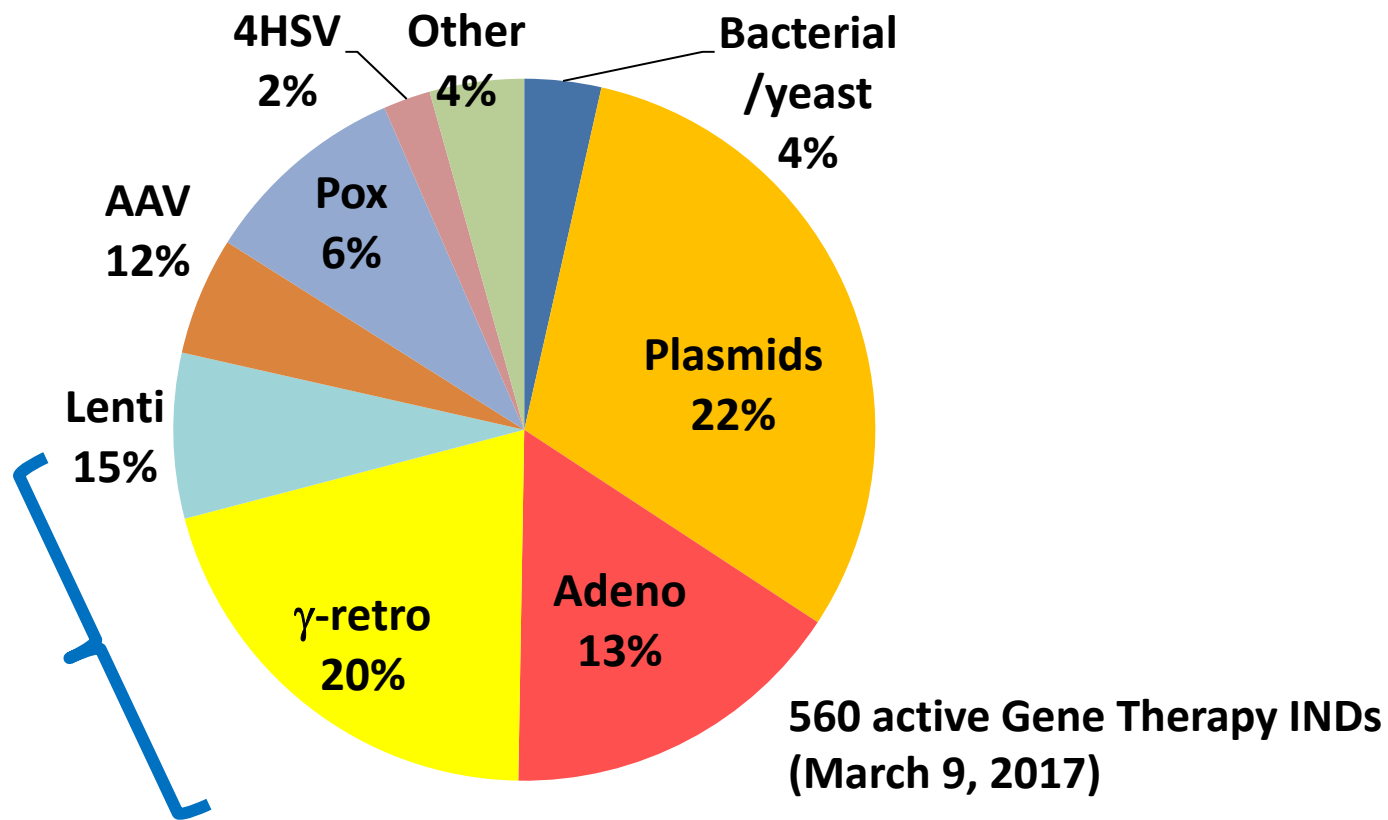


Safety Considerations for Gene Editing and Other Gene Therapy Products: An FDA Perspective

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Division of Cellular and Gene Therapies
Office of Tissues and Advanced Therapies
CBER, FDA

Gene therapy products regulated by OTAT



Clinical applications of lentiviral vectors

***Ex vivo* applications**

- T cells to treat HIV infection
- CAR T cells to treat leukemias and lymphomas
- CD34+ hematopoietic progenitor/stem cells to treat
 - blood disorders
 - metabolic disorders
 - immunodeficiencies

***In vivo* applications**

- Ocular diseases
- CNS disorders
- Immunotherapy applications



Safety issues with HIV-1-based lentiviral vectors used in clinical trials

Potential to form replication-competent lentivirus

Potential for insertional gene activation/inactivation

Potential for off-target transduction *in vivo*

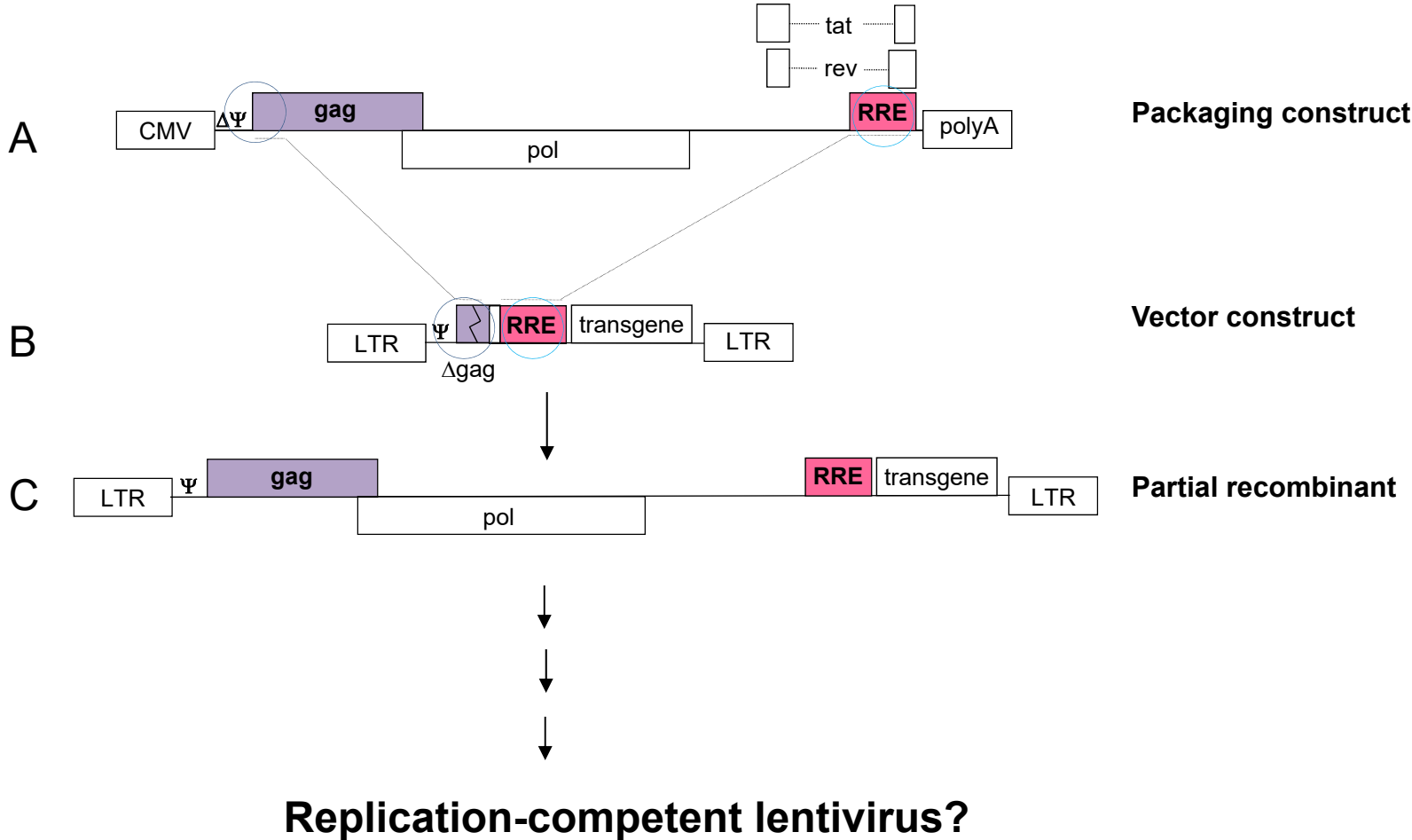
Approaches to improve the safety of lentiviral vectors

- Reducing the likelihood to form replication-competent lentivirus
- Narrowing the vector's tissue tropism
- Directing vector integration to genomic “safe harbor sites”
- Modifying the cell substrate for lentiviral vector production

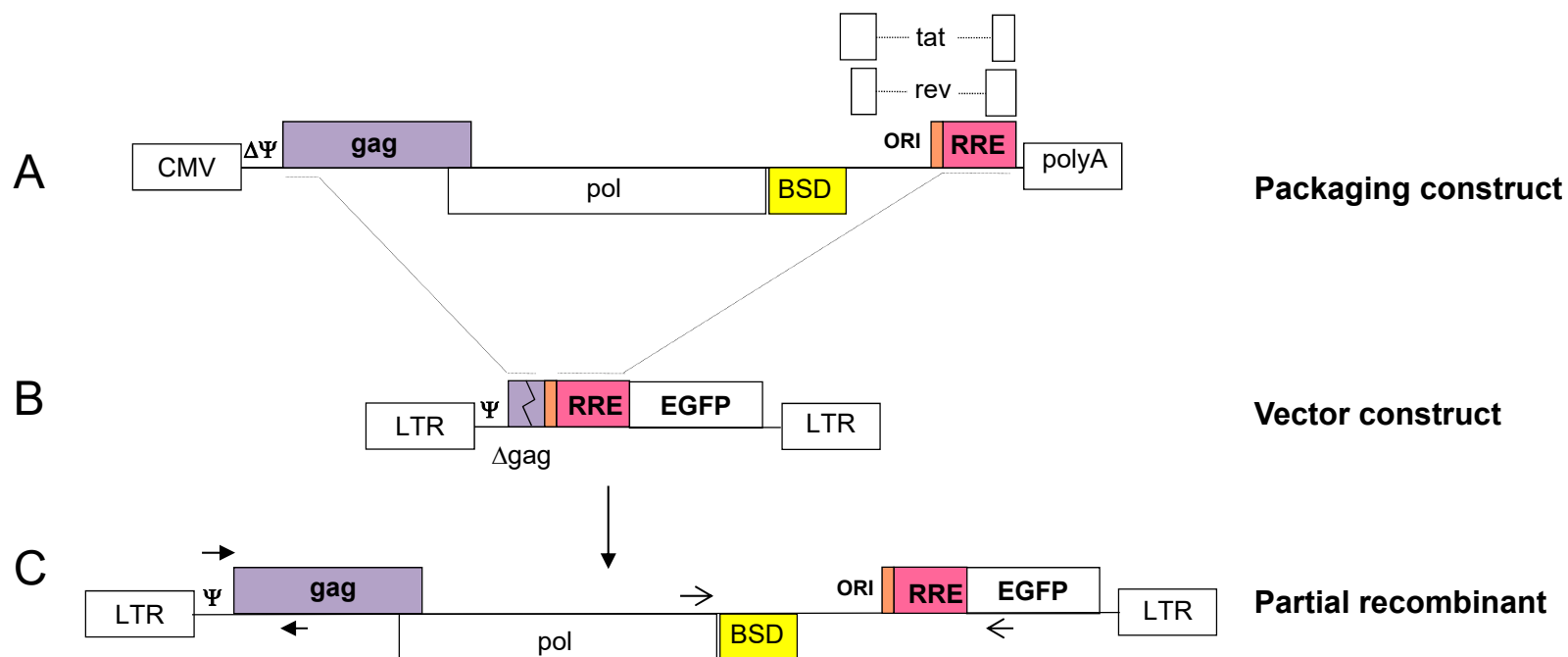
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Emergence of partial recombinants



Assay to assess formation of partial recombinants



Transduction of HEK 293 cells

Blasticidin (BSD) selection

Characterization of resistant cell clones by PCR and DNA sequencing

Reducing sequence overlaps

		Sequence overlap (%)	No. of BSD-resistant colonies per 10 ⁶ TU
A	<p>NL-EGFP(MSCV) vector</p>	13.3	63.4 ± 15.4
B	<p>NL-SRRE-EGFP(MSCV) vector</p>	9.8	25.6 ± 5.3
C	<p>NL-SRRE/ΔORI-EGFP(MSCV) vector</p>	9.1	9.5 ± 2.4
D	<p>NL(CMV)EGFP/CMV/WPREΔU3 vector</p>	15.6	3.2 ± 0.5



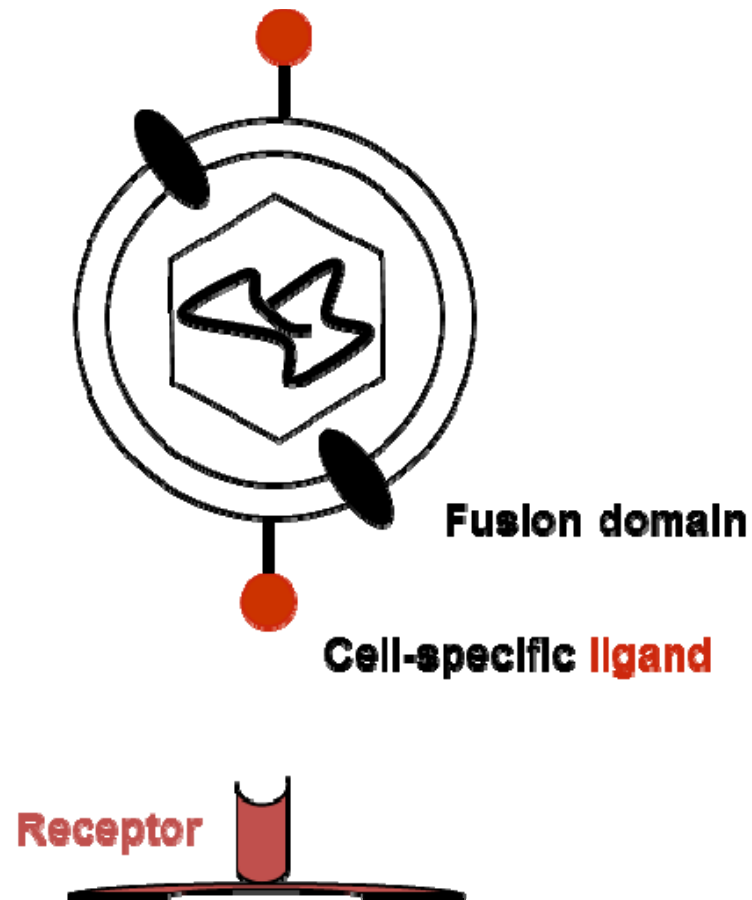
Conclusions

- Partial recombinants are rare
- The frequency of partial recombinants is dependent on vector design

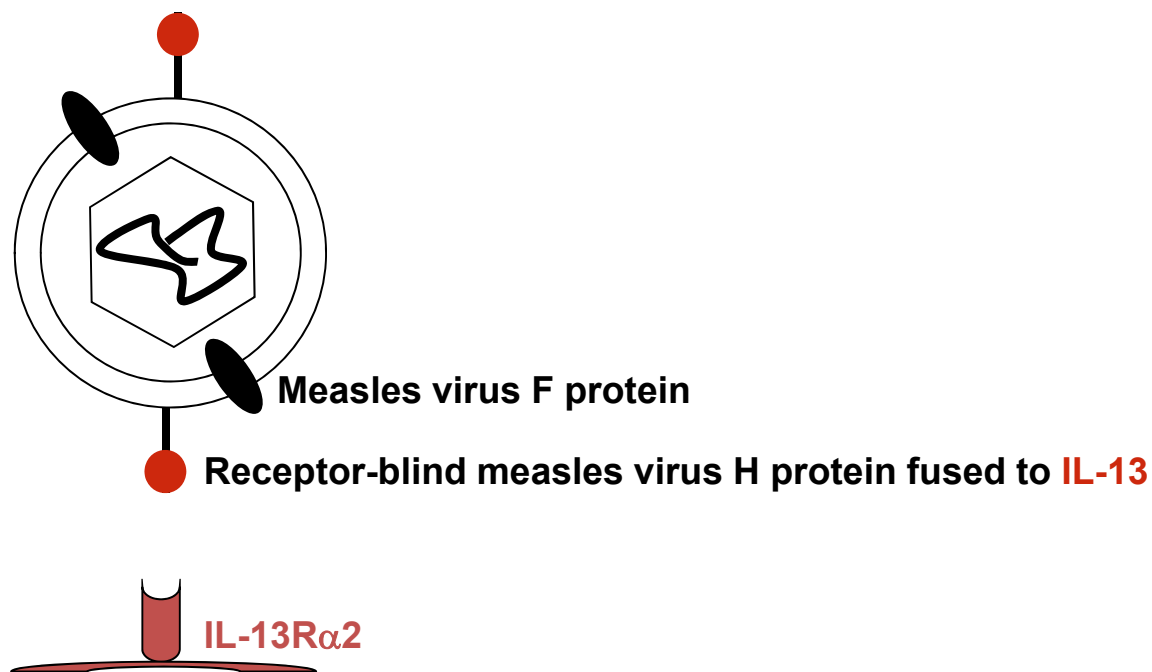
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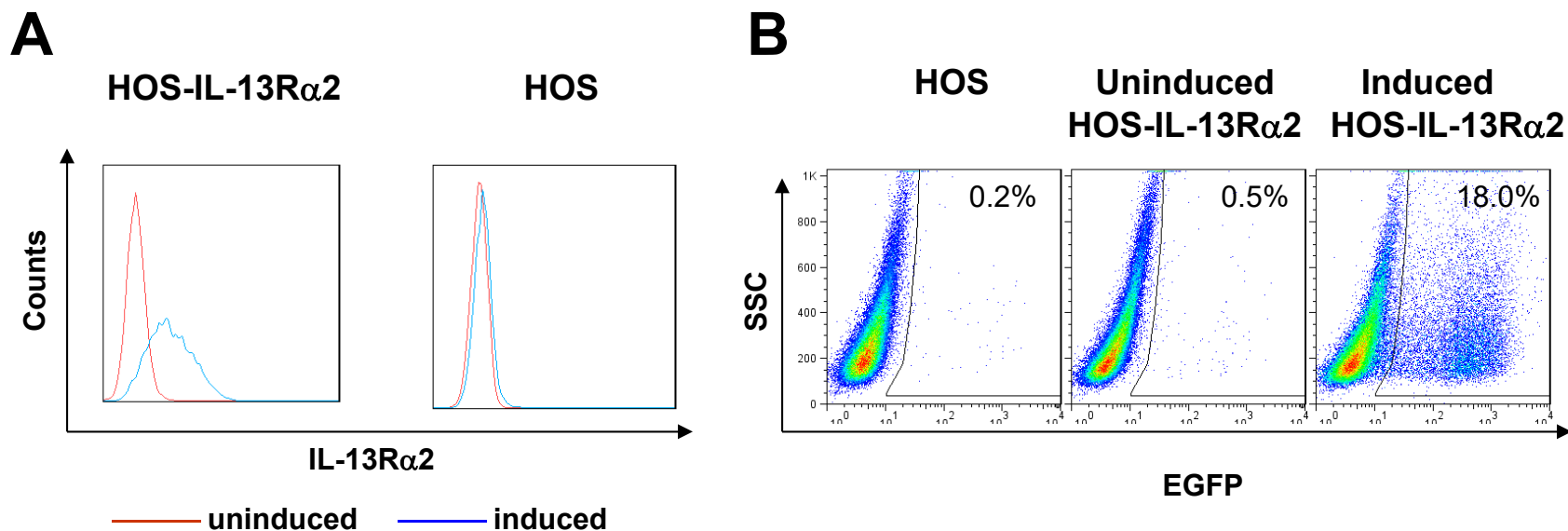
Strategy to target lentiviral vector transduction



Strategy to target IL-13R α 2-positive cells using lentiviral vectors pseudotyped with measles virus-derived hemagglutinin (H) and fusion (F) proteins

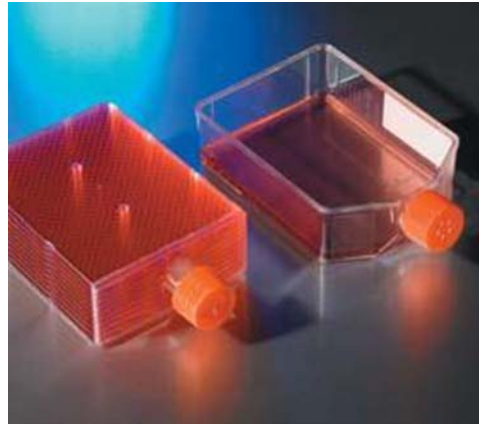


Targeting human osteosarcoma cells that conditionally overexpress IL-13R α 2



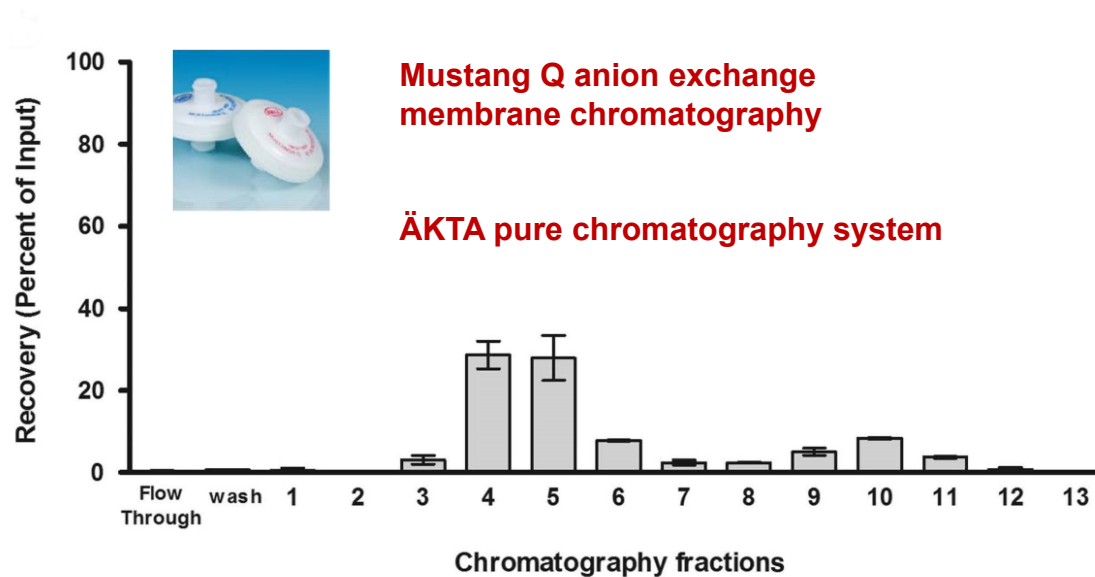
Ou et al. *Human Gene Therapy Methods* 23, 137-147 (2012)

Large-scale manufacturing of lentiviral vectors pseudotyped with measles virus H and F proteins



HYPERFlask Cell Culture Vessel

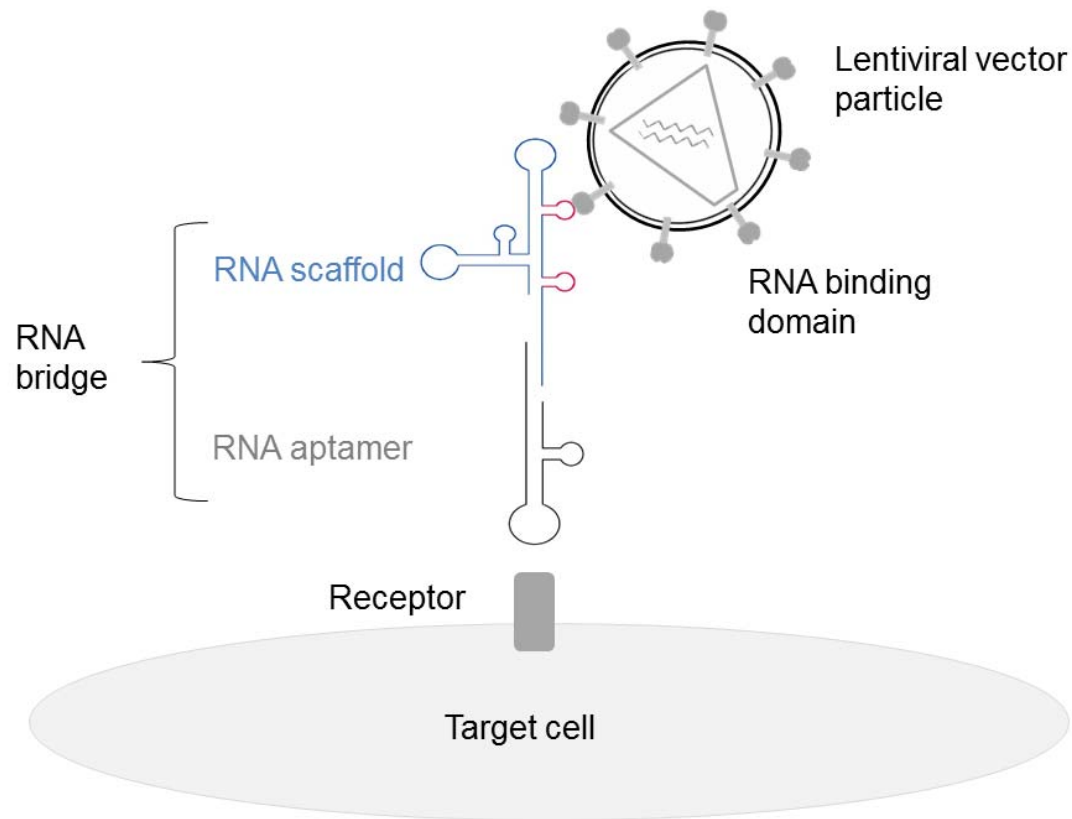
Large-scale manufacturing of lentiviral vectors pseudotyped with measles virus H and F proteins



Samples	Transducing units, TUs (%)	TUs/mg protein
Input vector samples	100	$2.43 \times 10^4 \pm 1.21 \times 10^4$
Vector eluate	64.36 ± 11.43	$2.87 \times 10^6 \pm 0.15 \times 10^6$
Samples after desalting using Amicon units	60.45 ± 27.64	$2.85 \times 10^6 \pm 0.33 \times 10^6$

100-fold purification

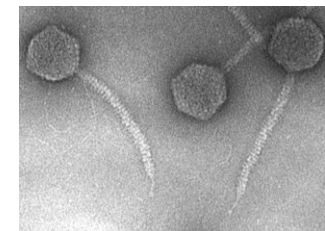
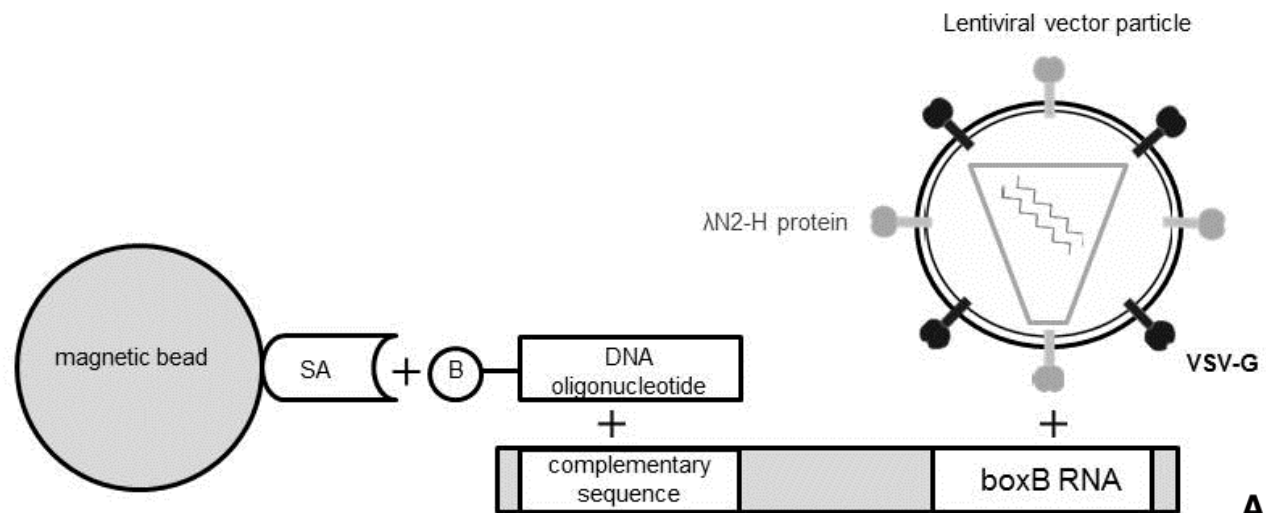
Depiction of a lentiviral vector targeting system involving RNA aptamers



Advantages of aptamer approach:

- RNA sequence space is large
- High affinity aptamer variants are available/can be obtained
- Ease of design/selection/production

Binding of boxB RNA to lentiviral vectors displaying an λ N2-H fusion protein

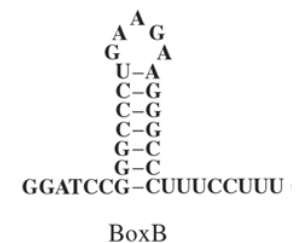


Bacteriophage λ

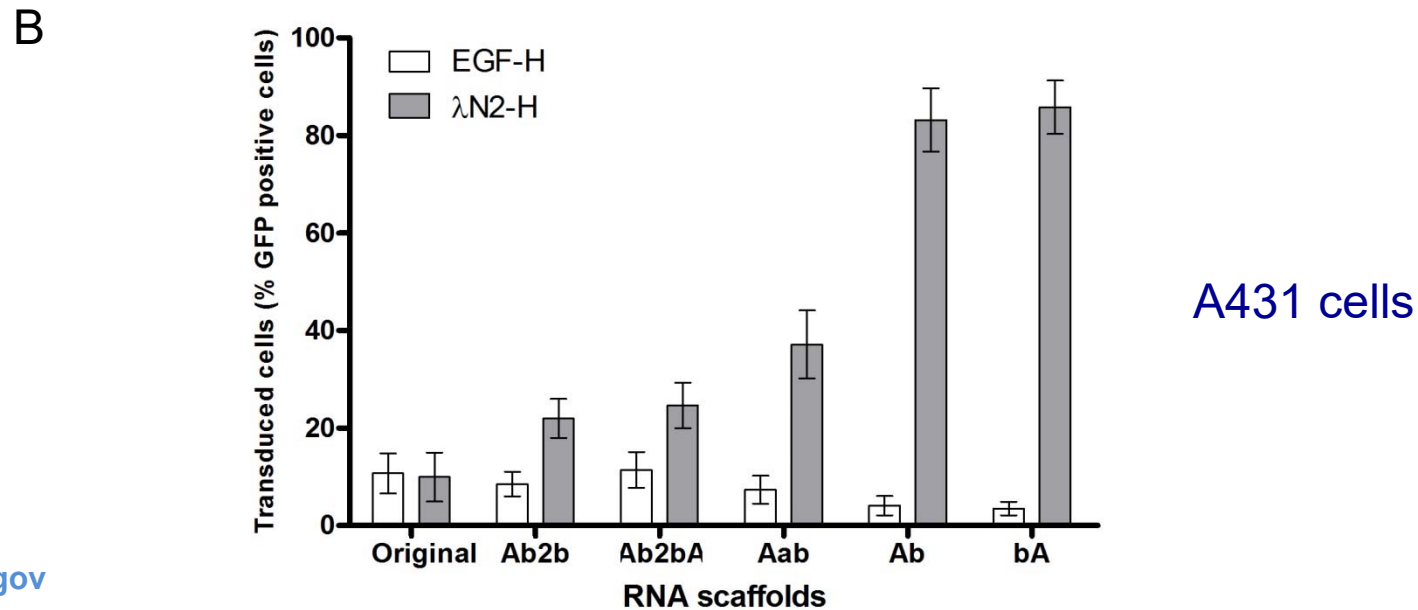
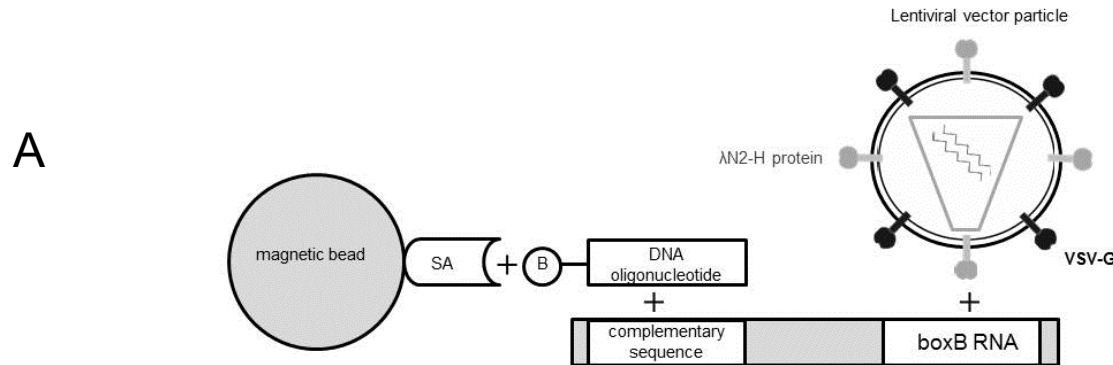
MDAQTRRRERRRAEKQAQWKAAN

A

B

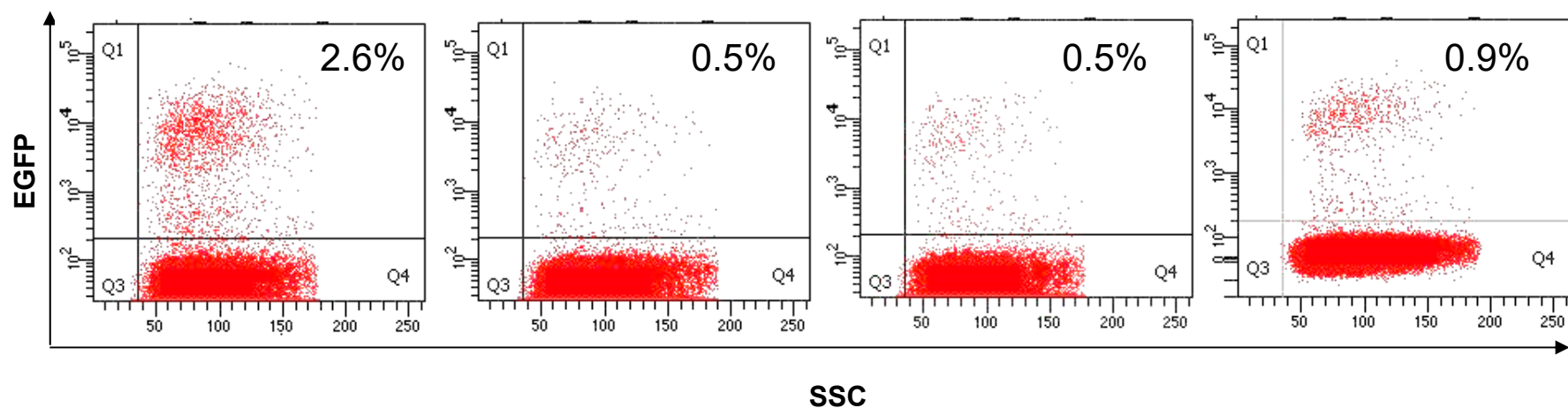


Binding of boxB RNA to lentiviral vectors displaying an λ N2-H fusion protein



Transduction of human epidermoid carcinoma A431 cells using lentiviral vector particles displaying an EGFR-specific RNA aptamer

H-λN	+	+	+	+
Scaffold	+	-	+	+
Aptamer	+	-	+	+
RNase	-	-	+	-
EGF	-	-	-	+



Conclusions

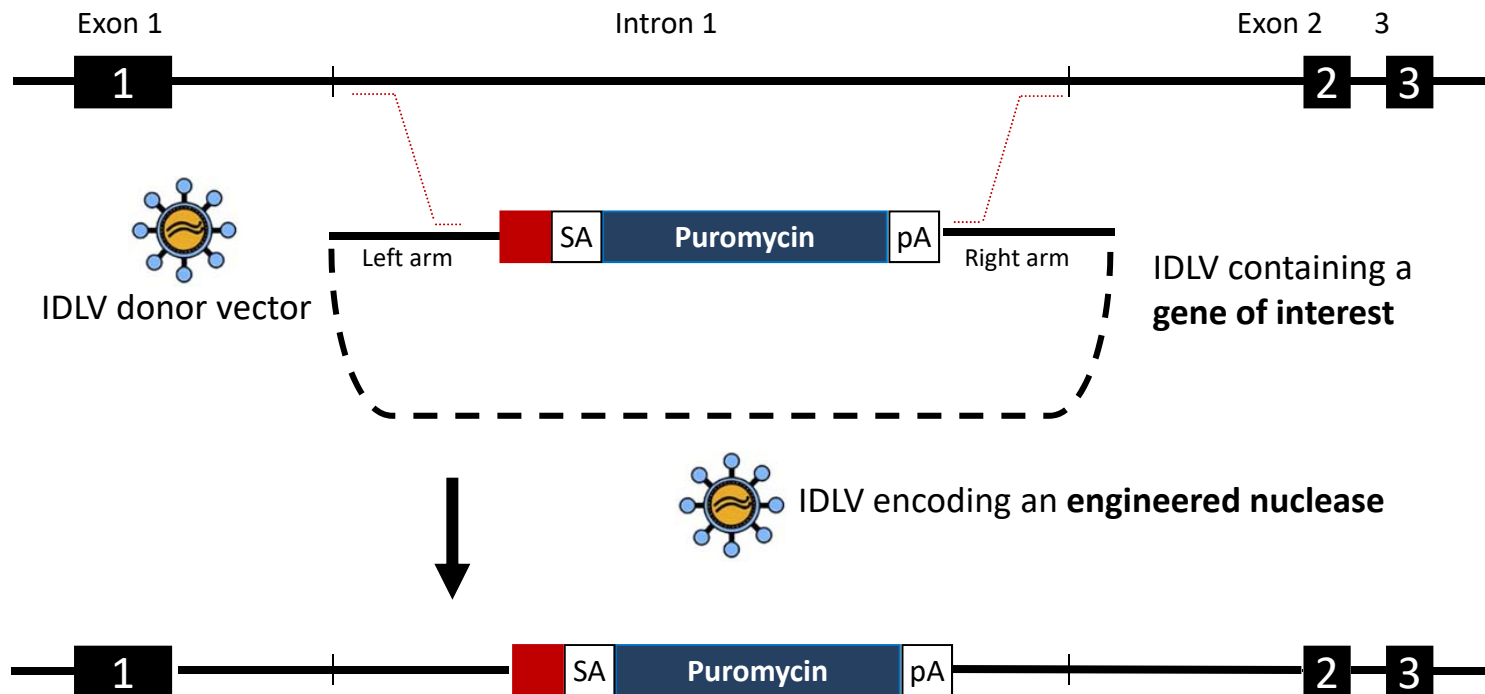
- Strategy involving measles virus H and F proteins allowed selective transduction of IL-13R α 2 positive cells
- Aptamer strategy looks promising but needs optimization

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Nuclease-mediated integration of integrase-defective lentiviral vectors (IDLVs) at the *AAVS1* “safe harbor” site on chromosome 19

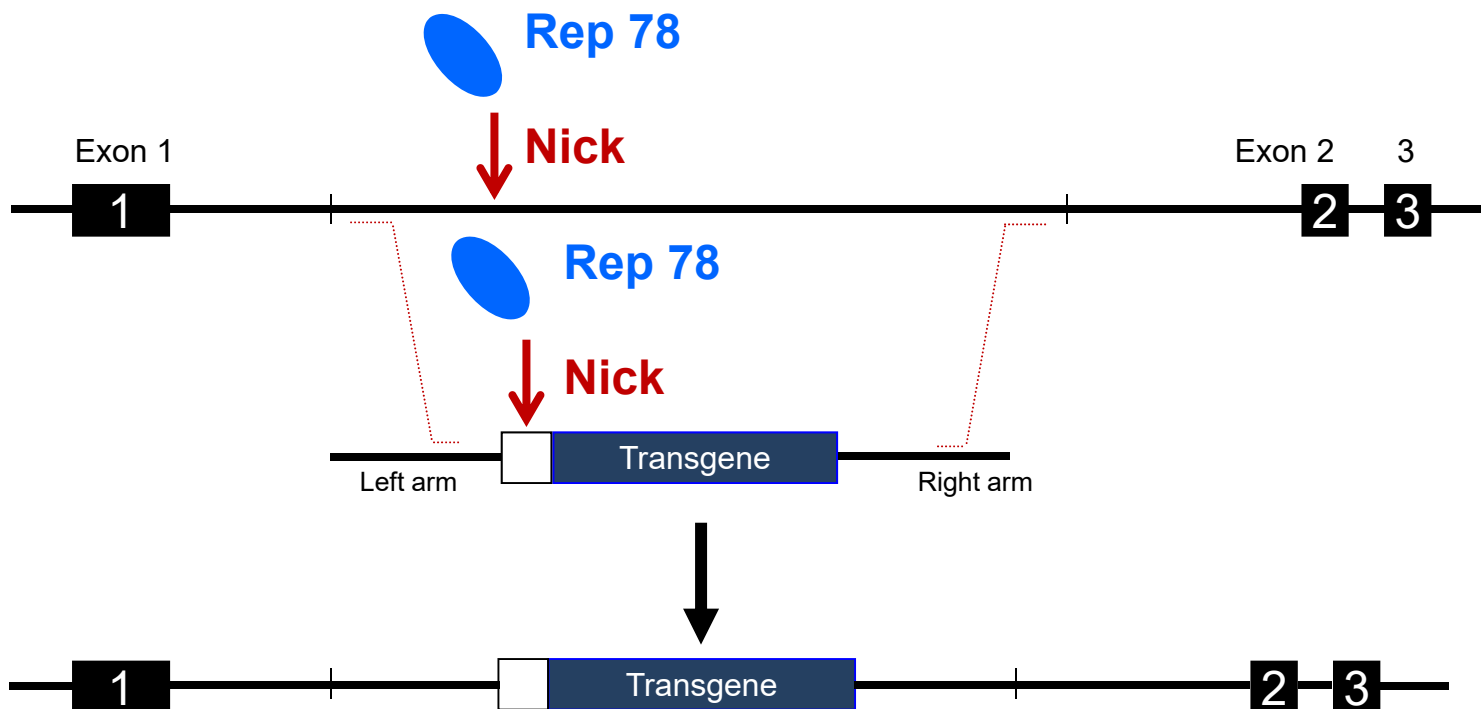
AAVS1 locus



Puromycin cassette integrated at *AAVS1* site

Double nick strategy involving the AAV2 Rep protein to promote “homologous recombination” at the AAVS1 locus

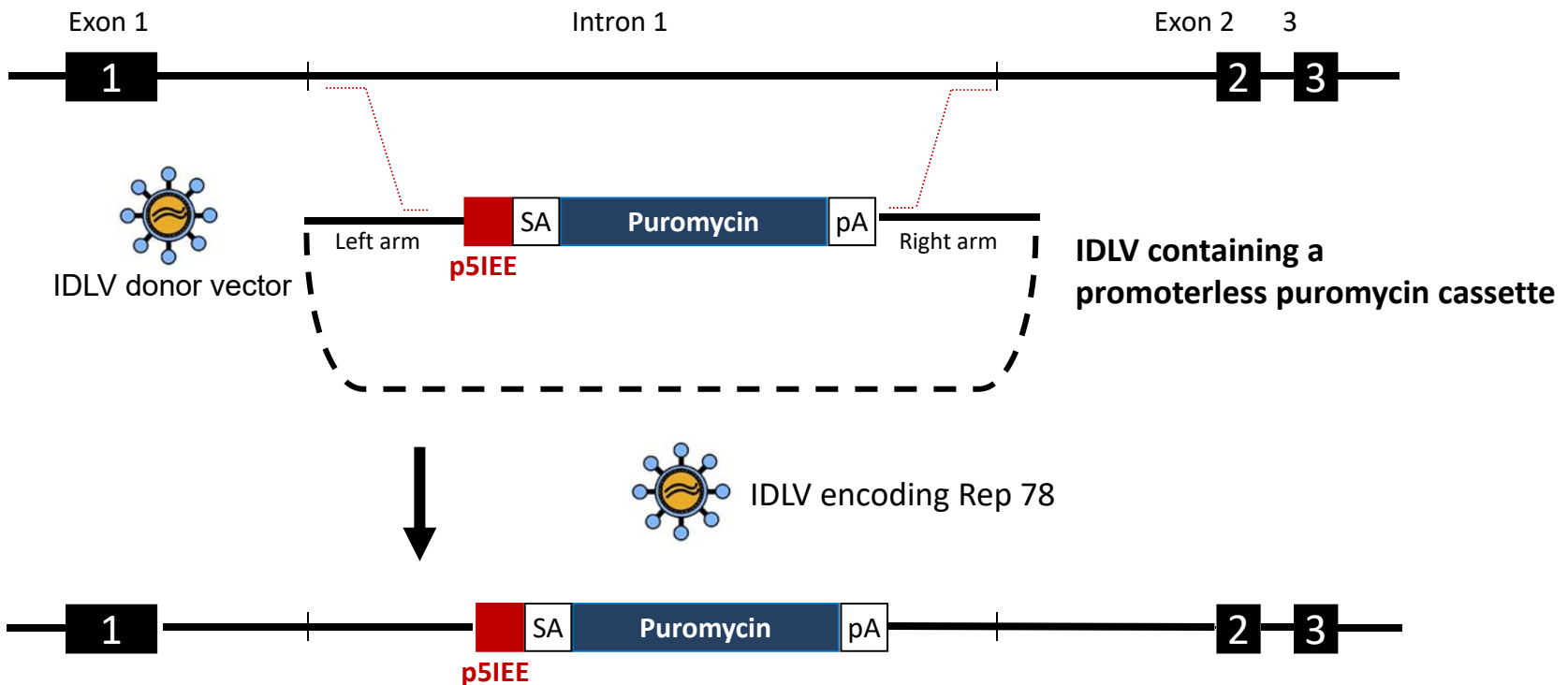
AAVS1 locus



Gonçalves MA et al. *Nucleic Acids Res.* 40, 3443-3455, 2012

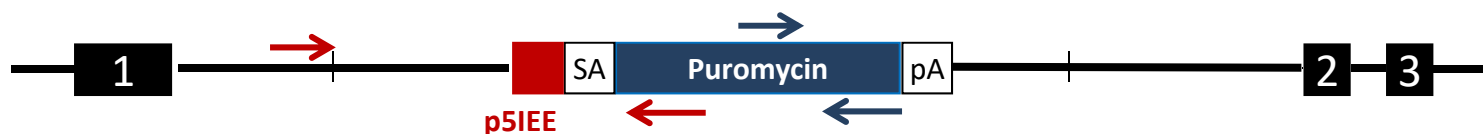
Rep-mediated integration of IDLVs at the AAVS1 “safe harbor” site

AAVS1 locus



Puromycin cassette integrated at AAVS1 site

Rep and ZFN-mediated insertion of transgene sequences at the AAVS1 site mediated by IDLVs



Samples	Number of puromycin-resistant clones*	PCR positive clones
Donor	66 ± 12	0% (0/10)
Donor + Rep	275 ± 14	84% (21/25)
Donor + ZFN	200 ± 10	100% (19/19)

*10⁵ cells used for transduction





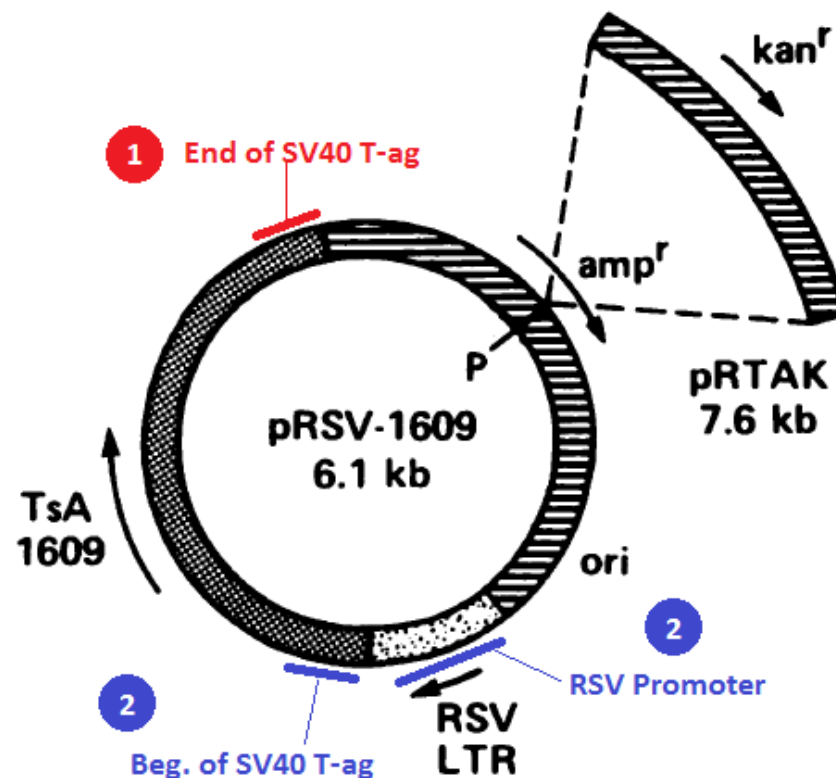
Conclusions

- The overall efficiencies of the nuclease/nickase approaches need to be improved
- ZFNs and Rep can be toxic if expressed at high levels

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Design of a HEK 293T cell line bearing a genomic deletion of the SV40 T antigen coding region

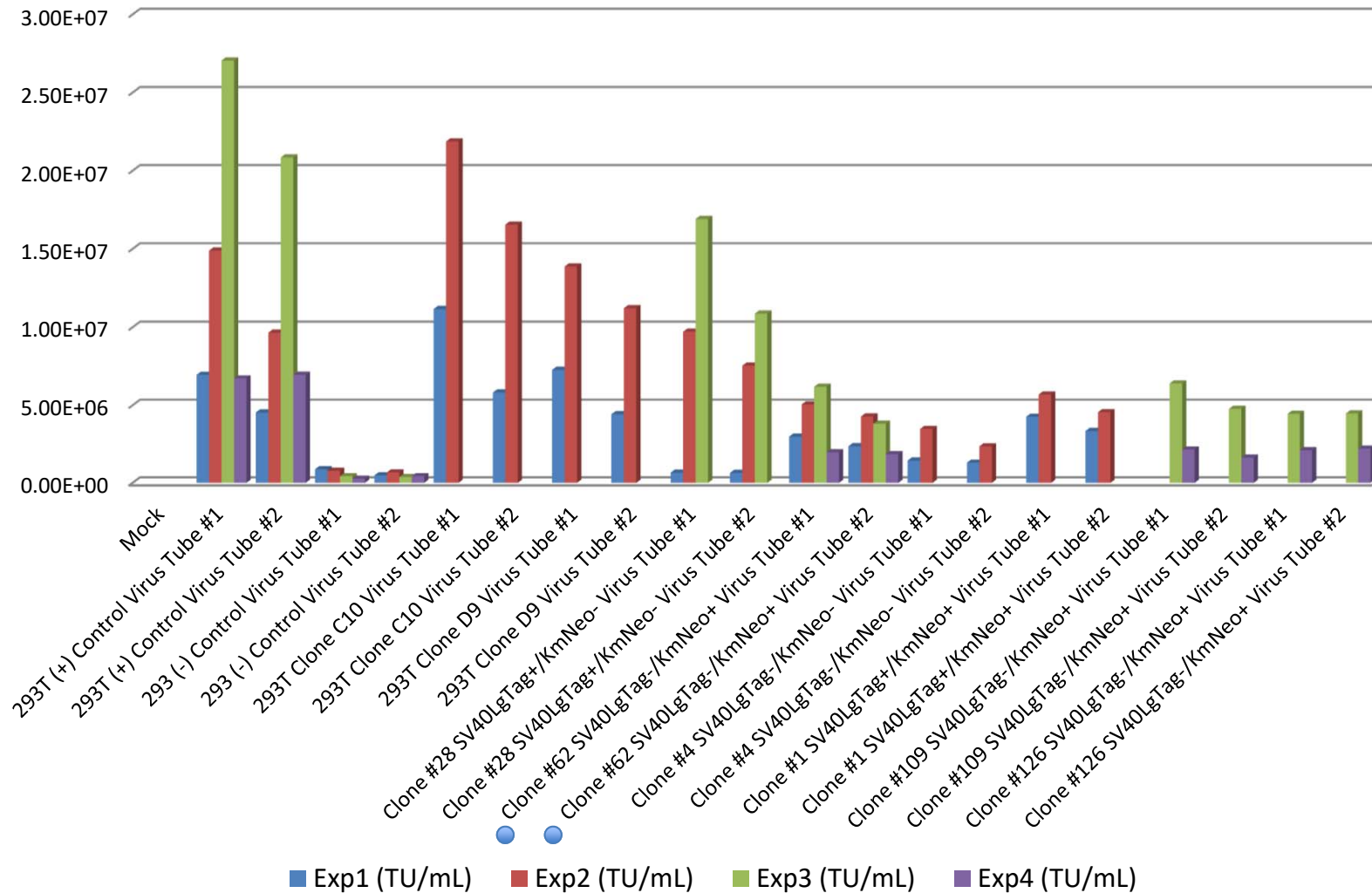


PCR screens



Total # of clones screened by PCR	176
● # of clones negative both for SV40 T-ag and Km/Neo resistance gene sequences	55
● # of clones negative for Km/Neo sequence	37
● # of clones negative PCR result for SV40 T-ag sequence	3 Clone #62, Clone #109, Clone #126
● # of wild type clones	81

Lentiviral vector production using modified HEK 293T cell clones



Conclusions

- Three out of the 176 cell clones screened revealed deletions of the T antigen sequence
- Vector titers for the three T antigen knock out clones were lower than those obtained using HEK 293T cells but higher than those obtained using HEK 293 cells

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Questions?